

Neurotoxicity, Neuroplasticity, and Magnetic Resonance Imaging Morphometry

What Is Happening in the Schizophrenic Brain?

Daniel R. Weinberger, MD; Robert K. McClure, MD

In an era of dramatic discoveries in neuroscience and genetics, it is likely that many popular theories and formulations about mental illness will need to be revised, if not discarded. The “neurodevelopmental hypothesis” is one of the popular theories about the origins of schizophrenia, which posits that abnormalities of early brain development increase risk for the subsequent emergence of the clinical syndrome.¹⁻³ An early piece of evidence in support of this hypothesis was the apparent lack of progression of cerebral ventricular enlargement observed with computed tomography during illness.⁴⁻⁹ An important assumption of the neurodevelopmental hypothesis is that the putative primary pathologic condition of the brain is a reflection of abnormalities of early development. The neurodevelopmental hypothesis thus assumes that developmental neuropathologic conditions should arrest early in life and not continue to progress. The computed tomography results showing no apparent progression seemed consistent with this assumption. However, a recent series of magnetic resonance imaging (MRI) studies has called into question this assumption, by revealing changes in measurements of brain structures over short periods in patients who have been ill for varying durations and at various stages of life. These recent studies¹⁰⁻¹⁴ have generated enthusiasm for a “neurodegenerative hypothesis,” harkening back to proposals of Kraepelin and other neuropathologists during the first quarter of the 20th century that there is destruction of neural tissue associated with psychosis. In fact, results of MRI measurements have been cited as support for a much broader conceptual revolution in psychiatry, a “neurotoxicity hypothesis” for many psychiatric illnesses, including affective disorders^{15,16} and anxiety and stress disorders¹⁷⁻¹⁹ and even jet lag.²⁰ This recent trend has been bolstered by basic discoveries about the adaptability of neuronal connections²¹ and the viability and reproducibility of neurons in the adult brain (eg, apoptosis and neurogenesis).^{22,23} These developments have led some to opine that the neurodegenerative hypothesis of schizophrenia may have been unjustly overshadowed by the ascendancy of the neurodevelopmental hypothesis.²⁴

While we are uncertain what constitutes appropriate relations between these 2 hypotheses, we are concerned that enthusiasm for the notion of neurodegeneration in schizophrenia (and in other psychiatric disorders) has outpaced the strength of the evidence. Quantitative measurements of cerebrospinal fluid spaces and tissue volumes on an MRI scan cannot establish that the brain has developed

abnormally, nor can they establish that tissue has degenerated. If the principal evidence for abnormal brain development in schizophrenia rested on the interpretation of computed tomography or MRI volume measurements, this hypothesis would not deserve serious consideration. The evidence that abnormal brain development may be a risk factor for schizophrenia comes from several domains, including: (1) abnormalities of early motor and cognitive development and histories of

From the Clinical Brain Disorders Branch, Intramural Research Program, National Institute of Mental Health, National Institutes of Health, Bethesda, Md.

Studies Showing Morphometric Changes

| Investigators | Average Age of First Scan (years) | Average Length Follow Up (years) | Brain Region Showing Progressive Change | Maximum % Change (per Year) |
|---|-----------------------------------|----------------------------------|--|---|
| DeLisi, et al, 1992, 1995, 1997, and 1998 | 26.4 | 1, 4, 4.7 | Left Lateral Ventricle Left and Right Cerebral Hemispheres Right Cerebellum Corpus Callosum (Isthmus) | (+) 3.0 (-) 1.42 (Right) (-) 2.2 (-) 1.14 |
| Rapaport et al, 1997 | 14.8 | 2 | Total Cerebrum Lateral Ventricles and VBR (Left>Right) Caudate Globus Pallidus Thalamic Area | (-) 2.58 (+) 9.7 (-) 4.1 (-) 9.9 (-) 7.1 |
| Jacobsen et al, 1998 | 15.2 | 2 | Total Cerebrum Right Temporal Lobe Superior Temporal Gyrus (Total) Superior Temporal Gyrus (Posterior) Right Superior Temporal Gyrus (Anterior) Left Hippocampus | (-) 2.30 (-) 4.15 (-) 3.7 (-) 4.3 (-) 3.2 (-) 7.15 |
| Nair et al, 1997 | 31.3 | 2 | Lateral Ventricle | (+) 12.48 |
| Garver et al, 2000 | 35.8 | 2.6 | None | |
| Gur et al, 1998 | 29.2 | 2.5 | Frontal Lobe (Left>Right) Temporal Lobe (Left>Right) | (-) 4.2 (Left) (-) 3.4 (Left) |
| Davis et al, 1998 | 39.5 | 5.1 | VBR (Left>Right) in Kraepelinian patients | (+) 3.3 |
| Lieberman et al, 2001 | 26 | 1.5 | Cerebral cortex in poor outcome patients Total Ventricles in poor outcome patients | (-) 0.42 (females) (-) 5.4 (females) |
| Mathalon et al, 2001 | 39.4 | 3.3 | Prefrontal Sulci Right Prefrontal Gray Right Frontal Sulci Left Frontal Gray Posterior Superior Temporal Sulci Posterior Superior Temporal Gray Left Lateral Ventricle | (+) 6.63 (Left) (-) 2.12 (-) 2.71 (-) 1.72 (-) 9.65 (Left) (+) 3.35 (Right) (+) 12.96 |

obstetrical adversity, (2) absence of evidence of neurodegeneration in postmortem tissue studies, and (3) association of developmental pathologic conditions with adult emergence of psychosis and related phenomena in animal and neurological models.¹⁻³ This represents a substantial body of research relating developmental compromise to subsequent schizophrenia and substantiating the neurobiological plausibility of this formulation. However, all of this evidence would be moot if neurodegeneration were apparent in postmortem studies. In contrast to predictions of a neurodegenerative hypothesis, the schizophrenic brain typically does not show loss of cortical neurons, gliosis, or any consistent evidence of degenerating or degenerated neurons.²⁵⁻²⁷ It is often stated in the psychiatric literature that

these negative findings are consistent with apoptosis.^{13,14,24} The basis for this assertion is obscure, perhaps arising from research on apoptosis in early brain development or in non-central nervous system cancers (some of which are associated with cell death without inflammation) and from investigations of experimental excitotoxicity that were of short duration.²⁸

Although apoptosis in early brain development does not typically initiate molecular activation of glia (although it may²⁹), in most pathologic brain conditions associated with apoptosis—including epilepsy,³⁰ experimental excitotoxicity,^{31,32} normal aging,³³ alcohol toxicity,^{34,35} steroid toxicity,³⁶ Alzheimer disease,³⁷ and many others—astroglial activation is prominent.^{28,30,38-42} Moreover, in conditions associated with loss of

neurons and with apoptosis, the neurons and glia that survive and those that are dying increase expression of genes and proteins that are aimed at molecular compensation and restoration or are involved in cell suicide.^{43,44} These changes are characteristically not seen in brain tissue of patients with schizophrenia.^{43,45} Much has been made of the lack of gliosis in the schizophrenic brain as failing to support neurodegenerative hypotheses, but it is the lack of expression of genes involved in cellular responses to injury and to DNA fragmentation that most militates against neurotoxicity and neuronal destruction hypotheses. Furthermore, the sine qua non of apoptosis is cell death, and diminished populations of cortical neurons are generally not found in tissue, including that of older patients who have

**Brain Region Showing
No Change or Opposite Change**

Correlation with Clinical Change

Right and Left Temporal Lobes
Right and Left Superior Temporal Gyri
Right and Left Hippocampus/Amygdala
Right and Left Caudate
None

No significant correlation between clinical measures and changes in ventricle or cerebral hemisphere volume; overall, symptoms improved in patients

Overall, symptoms improved in patients

Left Temporal Lobe
Left Superior Temporal Gyrus (Anterior)
Left and Right Amygdala
Right Hippocampus

No significant correlation between temporal lobe volume decrease and hallucinations, delusions, or negative symptoms; overall, symptoms improved in patients

None

Not reported

Lateral ventricle
Total Brain

Worsening of symptoms correlated with decreased ventricular and increased total brain volume

Whole Brain
Cerebral Spinal Fluid Spaces

Improvement in most symptoms correlated with decreased frontal and temporal lobe volumes in previously treated patients; overall, symptoms improved in patients

None

No significant correlation between change in ventricle-to-brain ratio and negative symptoms in non-Kraepelinian patients

Caudate Nuclei in all patients
Hippocampus in all patients
Cerebral Cortex in all patients
Total Ventricles in all patients
Total Ventricles in good outcome patients
Cerebral Cortex in good outcome patients
Hippocampus in poor outcome patients

Total ventricle and cerebral cortex increase correlated with poor outcome; cerebral cortex and hippocampus increase correlated with good outcome

Left Prefrontal Gray
Left Frontal Sulci
Right Frontal Gray
Anterior Superior Temporal Sulci
Right Anterior Superior Temporal Gray

Overall, symptoms improved in patients

suffered psychotic symptoms all of their adult lives.^{25-27,46} Therefore, although the lack, to date, of neurodegenerative signs in postmortem tissue does not rule out tissue damage, it raises the bar for believing other evidence that neurodegeneration has taken place. At some point in the disquisition, for neurodegeneration to be a plausible scenario, sequelae of neurodegeneration must be observable at the tissue level.

The evidence said to support the notion of neurodegeneration, although not present at the tissue level, is based on 2 phenomena. First is the apparent progression of clinical aspects of the syndrome in some patients, such as personality deterioration, dilapidation, and treatment resistance.¹³ Curiously, in longitudinal studies of cognitive function (a potentially direct measure of the

integrity of cortical neuronal systems), most of the data do not support progression, at least during about the first 20 years of illness (reviewed in Rund⁴⁷). Many unfortunate human circumstances and behaviors appear to get worse over time in some individuals (eg, joblessness and homelessness), without necessarily implicating degeneration of brain tissue. Although unemployment for a long period may in fact be associated with dynamic changes in synaptic architecture—just as learning new behaviors and habits may involve changes in the connections made between cells—these presumably are plastic modifications (ie, potentially reversible), not toxic degenerations (which usually imply irreversibility).

The second line of evidence for neurodegeneration is progressive

changes in volume measurements in structural MRI studies. This literature is selectively abstracted in the **Table**. Although there are some studies^{24,48-56,58} that find evidence of changes in MRI volume measurements, the results vary considerably among these positive studies. Some changes, eg, in temporal lobes, frontal lobes, and cerebrospinal fluid spaces, have been found in more than 1 study. However, no 2 studies have found the same pattern of changes across all of these measures, and each study appears to have its own unique combination of results. Moreover, although correlations between changes in symptoms and in measurements have been reported in a few studies,^{51,52,54,55} these also vary, and, remarkably, in most studies, patients have improved symptomatically while their MRI changes have ap-

peared to progress. This is hardly what would be predicted as a result of progressive loss of brain tissue; it is difficult to conceive of neurodegeneration being associated with clinical improvement.

Further concerns about inferring neurodegeneration in schizophrenia from MRI volume measurements are raised by comparisons with neurological conditions in which neurodegeneration is established. The magnitude of the changes reported in the studies purporting to show neurodegeneration in schizophrenia is not trivial. For example, Mathalon et al²⁴ claim a 2% per year reduction in frontal gray matter in patients with schizophrenia who have been ill for a mean of 15 years before they were studied. This degree of change is near the range reported in Alzheimer disease for the hippocampus, a structure that is markedly degenerated in this illness.⁵⁹ Other investigations of schizophrenia have reported volume loss in the hippocampus of greater magnitude, as much as 7% per year,⁵⁴ yet postmortem studies of schizophrenia have had difficulty uncovering evidence of even slight hippocampal shrinkage.^{27,43} The mesial area of the thalamus was shrinking at the rate of 7% per year in a study⁵² of adolescent patients with already chronic disease. Presumably, the tissue loss would have to spontaneously arrest fairly soon, or these patients would be virtually thalamusless by middle age. In a recent study²⁴ of patients in their fifth decade of life, the left lateral ventricles were increasing at a rate of 13% per year, thus doubling in size every 8 years, again at a rate that is seen in Alzheimer disease.⁶⁰ In the study⁵² of adolescent patients who had been ill for several years, the rate of ventricular volume change was 10% per year, translating into a doubling period of 10 years. At this pace, by the time a patient with schizophrenia reached age 60 years, there would be little brain left. Although it has been countered that the progressive MRI changes in schizophrenia may not be linear and may not occur throughout the course of illness,^{11,52,54} similar progressive changes that have been reported in adolescent patients with several years of dis-

ease,^{52,60} first-episode patients in their 20s,⁵⁵ patients in their 40s,²⁴ and patients in their 50s⁵⁶ make this counterargument unconvincing (unless one concludes that the neurodegeneration only occurs during the few years when patients are part of a National Institute of Mental Health-funded study). It is hard to imagine how the magnitude and duration of changes observed in MRI studies of patients with schizophrenia could be occurring as a neurodegenerative process, whether by cell necrosis or apoptosis, without observable evidence of neuronal loss and other related changes in postmortem tissue.

If the MRI changes, which are found at least in some studies, do not reflect neurodegeneration, how might they be explained? Variations in MRI measurements can reflect differences in image acquisition and analysis techniques, alterations in neuronal and nonneuronal tissue compartments, physiological alterations in brain tissue (eg, changes in tissue perfusion, fat, and water content), and changes in other chemical constituents that make up living brain.⁶¹ Indeed, numerous studies have shown that changes in body weight,⁶² alcohol intake,^{63,64} steroid administration,⁶⁵ and hormonal status⁶⁶ can rapidly produce changes in MRI volume measurements, some of greater magnitude than those reported in the schizophrenia studies. Recent findings that ventricular size can alternatively increase, decrease, and then increase again in the same patients scanned repeatedly over a few months suggest that such changes may reflect physiological variations.^{51,57} A recent report⁶⁷ claimed that only 4 weeks of treatment with lithium carbonate increases cortical gray matter volume by 3%. Treatment with paroxetine hydrochloride was reported to decrease thalamic volume by 19% in 12 weeks.⁶⁸ Neuroleptics also may rapidly change brain volume,⁶⁹ perhaps because of changes in tissue perfusion, and many studies^{70,71} have shown a relationship between neuroleptic exposure and basal ganglia volume change.

It is also conceivable that, in addition to physiological variations, the MRI changes reflect some degree of neuroplastic adaptations to the environment or to the experience of being

psychotic, perhaps reflected in less abundant neuropil, as has been observed in postmortem tissue from patients with schizophrenia.²⁶ Indeed, investigations of gene and protein expression in schizophrenia suggest that there is decreased transcriptional drive related to various signal-processing pathways.⁴³ This may reflect environmental factors reducing the neuronal information-processing load. These factors may include unstimulating environments, the effect of long-standing diagnosis of a chronic disease, comorbid conditions (including smoking, alcohol or other drug use, and untreated medical problems), and effects of long-term medication use. It has become clear from studies in experimental animals that numerous environmental factors have an effect on neuronal plasticity and can be associated with regression of dendrites and spines, possibly reflecting reduced stimulus-linked gene expression. It is also conceivable that a brain that processes information abnormally, perhaps because of genetic and developmental aberrations that may be factors in schizophrenia, would make unique plastic adaptations to environmental experience. These nondegenerative adaptations, however, are potentially reversible (as part of how a brain does business with environmental stimuli), in contrast to the implications of changes that reflect neurodegeneration. Although the possibility of such plastic alterations in neuronal connectivity is intriguing, it is not clear that their magnitude would be observable at the level of an MRI measurement. In the often cited study⁷² of stress-associated cellular changes in the monkey hippocampus (one of the reports that launched the experiential neurotoxicity revolution), although dramatic cytoarchitectural changes were described, including loss of neurons and gliosis, there was no change in gross hippocampal volume! Moreover, in studies⁷³⁻⁷⁵ of patients with epilepsy, seizure frequency, a plausible measure of neurotoxicity load, tends not to correlate with hippocampal volume reductions seen on MRI.

Based on these various considerations, we caution investigators involved in MRI studies and readers of this literature to suspect the improbability of the reported volume changes as being related to neurodegenera-

tion. In our view, the implications for patients and families of such an extreme interpretation of the data require more definitive evidence, which cannot emerge from such MRI measurements.

Submitted for publication March 10, 2001; final revision received May 23, 2001; accepted September 28, 2001.

Corresponding author and reprints: Daniel R. Weinberger, MD, Clinical Brain Disorders Branch, Intramural Research Program, National Institute of Mental Health, National Institutes of Health, 10 Center Dr, Bldg 10, Room 3C-101, MSc 1255, Bethesda, MD 20892 (e-mail: weinberd@dirpc.nimh.nih.gov).

REFERENCES

- Weinberger DR. Implications of normal brain development for the pathogenesis of schizophrenia. *Arch Gen Psychiatry*. 1987;44:660-669.
- Marenco S, Weinberger DR. The neurodevelopmental hypothesis of schizophrenia: following a trail of evidence from cradle to grave. *Dev Psychopathol*. 2000;12:501-527.
- Cannon TD, Rosso IM, Bearden CE, Sanchez LE, Hadley T. A prospective cohort study of neurodevelopmental processes in the genesis and epigenesis of schizophrenia. *Dev Psychopathol*. 1999;11:467-485.
- Illowsky BP, Juliano DM, Bigelow LB, Weinberger DR. Stability of CT scan findings in schizophrenia: results of an 8 year follow-up study. *J Neurol Neurosurg Psychiatry*. 1988;51:209-213.
- Vita A, Sacchetti E, Valvassori G, Cazzullo CL. Brain morphology in schizophrenia: a 2- to 5-year CT scan follow-up study. *Acta Psychiatr Scand*. 1988;78:618-621.
- Hoffman WF, Ballard L, Turner EH, Casey DE. Three-year follow-up of older schizophrenics: extrapyramidal syndromes, psychiatric symptoms, and ventricular brain ratio. *Biol Psychiatry*. 1991;30:913-926.
- Jaskiw GE, Juliano DM, Goldberg TE, Hertzman M, Urow-Hamell E, Weinberger DR. Cerebral ventricular enlargement in schizophreniform disorder does not progress: a seven year follow-up study. *Schizophr Res*. 1994;14:23-28.
- Vita A, Giobbio GM, Dieci M, Garbarini M, Morganti C, Comazzi M, Invernizzi G. Stability of cerebral ventricular size from the appearance of the first psychotic symptoms to the later diagnosis of schizophrenia. *Biol Psychiatry*. 1994;35:960-962.
- Kemali D, Maj M, Galderisi S, Milici N, Salvati A. Ventricle-to-brain ratio in schizophrenia: a controlled follow-up study. *Biol Psychiatry*. 1989;26:756-759.
- Miller R. Schizophrenia as a progressive disorder: relations to EEG, CT, neuropathological and other evidence. *Progress Neurobiol*. 1989;33:17-44.
- DeLisi LE. Is schizophrenia a lifetime disorder of brain plasticity, growth and aging? *Schizophr Res*. 1997;23:119-129.
- Woods BT. Is schizophrenia a progressive neurodevelopmental disorder? toward a unitary pathogenetic mechanism. *Am J Psychiatry*. 1998;155:1661-1670.
- Lieberman JA. Is schizophrenia a neurodegenerative disorder? a clinical and neurobiological perspective. *Biol Psychiatry*. 1999;46:729-739.
- Olney JW, Farber NB. Glutamate receptor dysfunction and schizophrenia. *Arch Gen Psychiatry*. 1995;52:998-1007.
- Sapolsky R. The possibility of neurotoxicity in the hippocampus in major depression: a primer on neuron death. *Biol Psychiatry*. 2000;48:755-765.
- Bremner JD, Narayan M, Anderson ER, Staib LH, Miller HL, Charney DS. Hippocampal volume reduction in major depression. *Am J Psychiatry*. 2000;157:115-118.
- Bremner JD. Does stress damage the brain? *Biol Psychiatry*. 1999;45:797-805.
- McEwen BS. Stress, adaptation, and disease: allostasis and allostatic load. *Ann N Y Acad Sci*. 1998;840:33-44.
- Cho K. Chronic "jet lag" produces temporal lobe atrophy and spatial cognitive deficits. *Nat Neurosci*. 2001;4:567-568.
- Duman RS, Heninger GR, Nestler EJ. A molecular and cellular theory of depression. *Arch Gen Psychiatry*. 1997;54:597-606.
- Malenka RC, Nicoll RA. Long-term potentiation: a decade of progress? *Science*. 1999;285:1870-1874.
- Kempermann G, Kuhn HG, Gage FH. More hippocampal neurons in adult mice living in an enriched environment. *Nature*. 1997;386:493-495.
- van Praag H, Kempermann G, Gage FH. Running increases cell proliferation and neurogenesis in the adult mouse dentate gyrus. *Nat Neurosci*. 1999;2:266-270.
- Mathalon DH, Sullivan EV, Lim KO, Pfefferbaum A. Progressive brain volume changes and the clinical course of schizophrenia in men: a longitudinal magnetic resonance imaging study. *Arch Gen Psychiatry*. 2001;58:148-157.
- Pakkenberg B. Total nerve cell number in neocortex in chronic schizophrenics and controls estimated using optical disectors. *Biol Psychiatry*. 1993;34:768-772.
- Selemon LD, Rajkowska G, Goldman-Rakic PS. Elevated neuronal density in prefrontal area 46 in brains from schizophrenic patients: application of a three-dimensional, stereologic counting method. *J Comp Neurol*. 1998;392:402-412.
- Harrison PJ. The neuropathology of schizophrenia: a critical review of the data and their interpretation. *Brain*. 1999;122:593-624.
- Fix AS, Horn JW, Wightman KA, Johnson CA, Long GG, Storts RW, Farber N, Wozniak DF, Olney JW. Neuronal vacuolization and necrosis induced by the noncompetitive N-methyl-D-aspartate (NMDA) antagonist MK(+)-801 (dizocilpine maleate): a light and electron microscopic evaluation of the rat retrosplenial cortex. *Exp Neurol*. 1993;123:204-215.
- Lafarga M, Andres MA, Calle E, Berciano MT. Reactive gliosis of immature Bergmann glia and microglial cell activation in response to cell death of granule cell precursors induced by methylazoxymethanol treatment in developing rat cerebellum. *Anat Embryol (Berl)*. 1998;198:111-122.
- Honavar M, Meldrum BS. Epilepsy. In: Graham D, Lantos PL, eds. *Greenfield's Neuropathology*. New York, NY: Oxford University Press; 1997:931-971.
- Pollard H, Charriaud-Marlangue C, Cantagrel S, Represa A, Robain O, Moreau J, Ben-Ari Y. Kainate-induced apoptotic cell death in hippocampal neurons. *Neuroscience*. 1994;63:7-18.
- Filipkowski RK, Hetman M, Kaminska B, Kaczmarek L. DNA fragmentation in rat brain after intraperitoneal administration of kainate. *Neuroreport*. 1994;5:1538-1540.
- Higami Y, Shimokawa I. Apoptosis in the aging process. *Cell Tissue Res*. 2000;301:125-132.
- Freund G. Apoptosis and gene expression: perspectives on alcohol-induced brain damage. *Alcohol*. 1994;11:385-387.
- Brooks PJ. Brain atrophy and neuronal loss in alcoholism: a role for DNA damage? *Neurochem Int*. 2000;37:403-412.
- Haynes LE, Griffiths MR, Hyde RE, Barber DJ, Mitchell JJ. Dexamethasone induces limited apoptosis and extensive sublethal damage to specific subregions of the striatum and hippocampus: implications for mood disorders. *Neuroscience*. 2001;104:57-69.
- Behl C. Apoptosis and Alzheimer's disease. *J Neural Transm*. 2000;107:1325-1344.
- Zagulska-Szymczak S, Filipkowski RK, Kaczmarek L. Kainate-induced genes in the hippocampus: lessons from expression patterns. *Neurochem Int*. 2001;38:485-501.
- Sapolsky RM, Krey LC, McEwen BS. Prolonged glucocorticoid exposure reduces hippocampal neuron number: implications for aging. *J Neurosci*. 1985;5:1222-1227.
- Carrasco J, Penkowa M, Hadberg H, Molinero A, Hidalgo J. Enhanced seizures and hippocampal neurodegeneration following kainic acid-induced seizures in metallothionein-I + II-deficient mice. *Eur J Neurosci*. 2000;12:2311-2322.
- LaFerla FM, Tinkle BT, Bieberich CJ, Haudenschild CC, Jay G. The Alzheimer's A beta peptide induces neurodegeneration and apoptotic cell death in transgenic mice. *Nat Genet*. 1995;9:21-30.
- Ferrer I, Blanco R. N-myc and c-myc expression in Alzheimer's disease, Huntington disease and Parkinson disease. *Brain Res Mol Brain Res*. 2000;77:270-276.
- Weinberger DR. Cell biology of the hippocampal formation in schizophrenia. *Biol Psychiatry*. 1999;45:395-402.
- Adams JM, Cory S. The Bcl-2 protein family: arbiters of cell survival. *Science*. 1998;281:1322-1326.
- Jarskog L, Gilmore JH, Selinger ES, Lieberman JA. Cortical Bcl-2 protein expression and apoptotic regulation in schizophrenia. *Biol Psychiatry*. 2000;48:641-650.
- Arnold SE, Trojanowski JQ, Gur RE, Blackwell P, Han LY, Choi C. Absence of neurodegeneration and neural injury in the cerebral cortex in a sample of elderly patients with schizophrenia. *Arch Gen Psychiatry*. 1998;55:225-232.
- Rund BR. A review of longitudinal studies of cognitive functions in schizophrenia patients. *Schizophr Bull*. 1998;24:425-435.
- DeLisi LE, Hoff AL, Kushner M, Caley A, Stritzke P. Left ventricular enlargement associated with diagnostic outcome of schizophreniform disorder. *Biol Psychiatry*. 1992;32:199-201.
- DeLisi LE, Tew W, Xie S, Hoff AL, Sakuma M, Kushner M, Lee G, Shedlack K, Smith AM, Grimson R. A prospective follow-up study of brain morphology and cognition in first-episode schizophrenic patients: preliminary findings. *Biol Psychiatry*. 1995;38:349-360.
- DeLisi LE, Sakuma M, Tew W, Kushner M, Hoff AL, Grimson R. Schizophrenia as a chronic active brain process: a study of progressive brain structural change subsequent to the onset of schizophrenia. *Psychiatry Res*. 1997;74:129-140.

51. DeLisi LE, Sakuma M, Ge S, Kushner M. Association of brain structural change with heterogeneous course of schizophrenia from early childhood through five years subsequent to a first hospitalization. *Psychiatry Res*. 1998;84:75-88.
52. Rapoport JL, Giedd J, Kumra S, Jacobsen L, Smith A, Lee P, Nelson J, Hamburger S. Childhood-onset schizophrenia: progressive ventricular change during adolescence. *Arch Gen Psychiatry*. 1997;54:897-903.
53. Nair TR, Christensen JD, Kingsbury SJ, Kumar NG, Terry WM, Garver DL. Progression of cerebroventricular enlargement and the subtyping of schizophrenia. *Psychiatry Res*. 1997;74:141-150.
54. Jacobsen LK, Giedd JN, Castellanos FX, Vaituzis AC, Hamburger SD, Kumra S, Lenane MC, Rapoport JL. Progressive reduction of temporal lobe structures in childhood-onset schizophrenia. *Am J Psychiatry*. 1998;155:678-685.
55. Gur RE, Cowell P, Turetsky BI, Gallacher F, Cannon T, Bilker W, Gur RC. A follow-up magnetic resonance imaging study of schizophrenia: relationship of neuroanatomical changes to clinical and neurobehavioral measures. *Arch Gen Psychiatry*. 1998;55:145-152.
56. Davis KL, Buchsbaum MS, Shihabuddin L, Spiegel-Cohen J, Metzger M, Frecska E, Keefe RS, Powchick P. Ventricular enlargement in poor-outcome schizophrenia. *Biol Psychiatry*. 1998;43:783-793.
57. Garver DL, Nair TR, Christensen JD, Holcomb JA, Kingsbury SL. Brain and ventricular instability during psychotic episodes of the schizophrenias. *Schizophr Res*. 2000;44:11-23.
58. Lieberman J, Chakos M, Wu H, Alvir J, Hoffman E, Robinson D, Bilder R. Longitudinal study of brain morphology in first episode schizophrenia. *Biol Psychiatry*. 2001;49:487-499.
59. Laakso MP, Lehtovirta M, Partanen K, Riekkinen PJ, Soininen H. Hippocampus in Alzheimer's disease: a 3-year follow-up MRI study. *Biol Psychiatry*. 2000;47:557-561.
60. DeCarli C, Kaye JA, Horwitz B, Rapoport SI. Critical analysis of the use of computer-assisted transverse axial tomography to study human brain in aging and dementia of the Alzheimer type. *Neurology*. 1990;40:872-883.
61. Hajnal JV, Saeed N, Oatridge A, Williams EJ, Young IR, Bydder GM. Detection of subtle brain changes using subvoxel registration and subtraction of serial MR images. *J Comput Assist Tomogr*. 1995;19:677-691.
62. Swayze VW II, Andersen A, Arndt S, Rajarethinam R, Fleming F, Sato Y, Andreasen NC. Reversibility of brain tissue loss in anorexia nervosa assessed with a computerized Talairach 3-D proportional grid. *Psychol Med*. 1996;26:381-390.
63. Schroth G, Naegele T, Klose U, Mann K, Petersen D. Reversible brain shrinkage in abstinent alcoholics, measured by MRI. *Neuroradiology*. 1988;30:385-389.
64. Pfefferbaum A, Sullivan EV, Mathalon DH, Shear PK, Rosenbloom MJ, Lim KO. Longitudinal changes in magnetic resonance imaging brain volumes in abstinent and relapsed alcoholics. *Alcohol Clin Exp Res*. 1995;19:1177-1191.
65. Gordon N. Apparent cerebral atrophy in patients on treatment with steroids. *Dev Med Child Neurol*. 1980;22:502-506.
66. Denton ER, Holden M, Christ E, Jarosz JM, Russell-Jones D, Goodey J, Cox TC, Hill DL. The identification of cerebral volume changes in treated growth hormone-deficient adults using serial 3D MR image processing. *J Comput Assist Tomogr*. 2000;24:139-145.
67. Moore GJ, Bebchuk JM, Wilds IB, Chen G, Manji HK. Lithium-induced increase in human brain grey matter. *Lancet*. 2000;356:1241-1242.
68. Gilbert AR, Moore GJ, Keshavan MS, Paulson LA, Narula V, Mac Master FP, Stewart CM, Rosenberg DR. Decrease in thalamic volumes of pediatric patients with obsessive-compulsive disorder who are taking paroxetine. *Arch Gen Psychiatry*. 2000;57:449-456.
69. Scheepers FE, de Wied CC, Pol HE, van de Flier W, van der Linden JA, Kahn RS. The effect of clozapine on caudate nucleus volume in schizophrenic patients previously treated with typical antipsychotics. *Neuropsychopharmacology*. 2001;24:47-54.
70. Chakos MH, Lieberman JA, Bilder RM, Borenstein M, Lerner G, Bogerts B, Wu H, Kinon B, Ashtari M. Increase in caudate nuclei volumes of first-episode schizophrenic patients taking antipsychotic drugs. *Am J Psychiatry*. 1994;151:1430-1436.
71. Corson PW, Nopoulos P, Miller DD, Arndt S, Andreasen NC. Change in basal ganglia volume over 2 years in patients with schizophrenia: typical versus atypical neuroleptics. *Am J Psychiatry*. 1999;156:1200-1204.
72. Sapolsky RM, Uno H, Rebert CS, Finch CE. Hippocampal damage associated with prolonged glucocorticoid exposure in primates. *J Neurosci*. 1990;10:2897-2902.
73. Trenerry MR, Jack CR Jr, Sharbrough FW, Cascino GD, Hirschorn KA, Marsh WR, Kelly PJ, Meyer FB. Quantitative MRI hippocampal volumes: association with onset and duration of epilepsy, and febrile convulsions in temporal lobectomy patients. *Epilepsy Res*. 1993;15:247-252.
74. Cendes F, Andermann F, Gloor P, Lopes-Cendes I, Andermann E, Melanson D, Jones-Gotman M, Robitaille Y, Evans A, Peters T. Atrophy of mesial structures in patients with temporal lobe epilepsy: cause or consequence of repeated seizures? *Ann Neurol*. 1993;34:795-801.
75. Tasch E, Cendes F, Li LM, Dubeau F, Andermann F, Arnold DL. Neuroimaging evidence of progressive neuronal loss and dysfunction in temporal lobe epilepsy. *Ann Neurol*. 1999;45:568-576.